



REVIEW ARTICLE

Harnessing the genome for characterization of G-protein coupled receptors in cancer pathogenesis

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G-protein coupled receptors (GPCRs) mediate numerous physiological processes and represent the targets for a vast array of therapeutics for diseases ranging from depression to hypertension to reflux. Despite the recognition that GPCRs can act as oncogenes and tumour suppressors by regulating oncogenic signalling networks, few drugs targeting GPCRs are utilized in cancer therapy. Recent large-scale genome-wide analyses of multiple human tumours have uncovered novel GPCRs altered in cancer. However, work aiming to determine which GPCRs from these lists are the drivers of tumourigenesis, and hence valid therapeutic targets, comprises a formidable challenge. The present review highlights recent studies providing evidence that GPCRs are relevant targets for cancer therapy through their effects on known cancer signalling pathways, tumour progression, invasion and metastasis, and the microenvironment. Furthermore, the review also explores how genomic analysis is beginning to highlight GPCRs as therapeutic targets in the age of personalized medicine.

Introduction

G-protein coupled receptors (GPCRs) transduce extracellular signals from a variety of ligands through the plasma membrane, resulting in the modulation of intracellular signalling pathways [1]. This is accomplished in large measure by the activation of heterotrimeric G-proteins and downstream second messengers. Composed of over 900 members in humans, GPCRs are seven-transmembrane proteins that regulate many physiological processes, including vision, olfaction, taste, behaviour and autonomic nervous system transmission. This wide array of functions has resulted in the extensive utilization of GPCR targeted therapeutics, accounting for 30-50% of all currently used drugs, both prescription and over-the-counter. The wide use of GPCR drugs can also be attributed to GPCR localization on the cell surface, abrogating the

requirement for a drug to be cell permeable, as well as the ability of GPCRs to bind a variety of ligands, including antibodies, peptides and small molecules. Furthermore, GPCR signalling can be tightly regulated through the utilization of agonists, antagonists and inverse agonists. However, drugs targeting GPCRs are rarely utilized in cancer treatment, despite evidence that GPCRs mediate many aspects of tumourigenesis [2]. This evidence has been accumulating ever since the discovery in 1986 of the transforming activity of the GPCR mas [3]. More recently, genomic analyses have uncovered GPCR mutations, copy number alterations and gene expression and methylation changes in a wide variety of cancers [4-6]. Determining the biological implication of these genomic alterations will allow utilization of GPCR targeted therapeutics in those

Abbreviations

EGF, epidermal growth factor; EGFR, epidermal growth factor receptor; ER, oestrogen receptor; GPCR, G-protein coupled receptor; GPER, G-protein coupled oestrogen receptor; GRM, glutamate receptor, metabotropic; NSCLC, non-small-cell lung cancer; PAR, protease-activated receptor; PI3K, phosphoinositide 3-kinase; STAT3, signal transducer and activator of transcription-3.

patients with GPCR-driven tumours. The present review first highlights recent genomic analyses of large-scale tumour sets and discusses how these studies are providing insight into GPCR-driven tumourigenesis. Subsequently, recent advances implicating GPCR signalling in various stages of tumourigenesis are discussed, including the regulation of known oncogenic signalling networks. Finally, the implications of this work on the future of personalized cancer medicine are discussed. Because several comprehensive reviews have described the varied roles for GPCRs in mitogenic signalling and cancer, the present review focusses on recent advances in the field [2,7].

Identification of GPCRs in large-scale human tumour analyses

Advances in sequencing technologies and bioinformatics tools have resulted in the recent explosion in genomic information from human tumour samples [8,9]. These data have been coupled with information on copy number alterations, gene expression and DNA methylation to provide a glimpse into the complex world of human cancer. However, merely providing lists of genomic alterations in a specific disease is only the first step in the process of translating this knowledge into cancer therapeutics. As a result of the inherent complexity of the data obtained from such analyses, it is critical to utilize computational methods to identify statistically significant alterations. One must then perform biological experiments in the proper model system to determine the oncogenic (i.e. cancer promoting) or tumour suppressive (i.e. cancer suppressing) activity of any candidate.

Because whole genome analyses can provide an overwhelming deluge of information, several methods for decreasing this complexity have been utilized. These include focused analysis of a specific gene family, cancer from a specific tissue of origin or specific genomic alteration, or combining data from multiple analyses to find commonalities. The efficacy of the gene family approach was displayed in a study that sequenced 734 GPCRs in 11 melanoma samples [10]. Mutations in glutamate receptor (GRM)3 (metabotropic) were found to be frequent in melanoma and subsequent functional analysis provided a mechanism of action (increased mitogen-activated protein kinase kinase phosphorylation) and possible therapeutic intervention (mitogen-activated protein kinase kinase inhibition). In addition to the obvious implications for patients with GRM3 mutations, this study highlights an important facet of GPCR biology with respect to cancer therapeutics: as mediators of diverse cell signalling pathways, the effect of mutations in GPCRs can often be overcome through inhibition of downstream signalling effectors. Therefore, although GPCRs are themselves good drug targets, knowledge of GPCR alterations may be quickly translated to the clinic, even in the absence of specific agonists or antagonists. Because GPCRs are known to be highly druggable targets, they are often included in screens designed to detect the 'druggable genome'. One such analysis was performed on 441 mixed primary tumours, looking for mutations in known cancer genes, protein kinases, E3 ligases, deubiquitinases, GPCRs and other enzymes [4]. Of the 157 GPCRs analyzed, mutations were detected in 13, including GRM1, GRM8 and brainspecific angiogenesis inhibitor 3 in lung cancer. This information was then coupled with copy number analysis to identify further putative cancer genes. The cancer relevance of GRM8 was reinforced in a study coupling whole-exome sequencing and high-throughput screening for the detection of candidate driver genes in endometrial cancer [11]. Candidate genes from the sequencing effort were targeted for knockdown in endometrial cancer cell lines, providing functional validation. A novel method for the unbiased detection of tumour suppressor genes, in which loss of heterozygosity and allele-specific detection were uncovered via exome and transcriptome sequencing, revealed 21 potential tumour suppressive GPCRs in two cancer cell lines [12]. Although no validation was performed, this type of analysis can provide a starting point for selection of candidates for biological analysis. In addition to genetic mutations, differential DNA methylation can drive tumourigenesis. A screen designed to identify genes whose DNA methylation-mediated silencing is required for cancer cell survival uncovered several GPCRs, including the purinergic receptor P2RY14 [6]. Re-expression of P2RY14 in colon cancer cell lines decreased cell viability, suggesting that this method can detect novel tumour suppressors.

Large-scale whole genome analyses, such as those undertaken by The Cancer Genome Atlas (http://cancergenome.nih.gov/) seek to provide a comprehensive map of genomic and epigenomic alterations in a number of cancers [5]. Mutations in GPCRs were found to be a frequent event in lung squamous cell carcinoma. Genotyping of single nucleotide polymorphisms among thousands of breast cancer samples has recently uncovered novel loci associated with increased risk [13,14]. Among these, parathyroid hormone 1 receptor was identified as a plausible candidate gene from a newly identified rick locus [13] and LGR6 was found to associate specifically with oestrogen receptor (ER)-negative breast cancer [14]. These datasets can be mined for

specific GPCRs and again provide a starting point to pick genes for functional validation in the appropriate model systems. A recent analysis has utilized this information to attempt to fully characterize the mutational landscape of GPCRs in human tumours [9]. As the use of targeted therapeutics and sequencing of tumours expands, cancer treatment may well focus not on the organ or origin but on the specific genetic alterations present in the tumour. Therefore, uncovering the presence, and biological relevance, of these mutations in distinct tumour types is of great value. Accordingly, it was noted that thyroid-stimulating hormone receptor, known to be frequently mutated in thyroid cancer, is also mutated in several other cancers, including those of the large intestine, lung and ovary [9]. This analysis also uncovered the unexpected finding that adhesion family GPCRs are frequently mutated in a variety of cancers. These receptors are important for interactions with neighbouring cells and the extracellular matrix. Importantly, because GPCRs are known to regulate processes such as cell migration and immune cell function, assays focusing solely on cell viability will fail to capture the full extent of GPCR involvement in cancer pathogenesis.

Novel bioinformatics tools are being developed to objectively prioritize gene lists for biological analysis. One such tool utilizes multiple parameters, including structural information, drug target homology and chemoinformatics to identify the most chemically tractable targets [15]. As a proof-of-concept, an analysis was performed on the Cancer Gene Census list, containing 488 genes implicated in human tumourigenesis, including several GPCRs. Thyroid-stimulating hormone receptor was identified as a putative druggable target in thyroid cancer and Smoothened (a Hedgehog pathway component) as a target in glioblastoma multiforme. Extension of this analysis to the entire GPCR family may identify novel drug targets, as well as expand the clinical utility of Food and Drug Association-approved therapeutics into new patient cohorts. These recent papers have clearly demonstrated the power of largescale genomic analysis on the identification of putative cancer-associated GPCRs. Many of them, however, lack the functional validation necessary for further clinical evaluation. Below, studies focused on biological target validation, a critical step in translating genomic information to the clinic, are highlighted.

GPCR regulation of cancer signalling pathways

Once a cancer-associated GPCR has been identified, it is critical to determine the underlying biology of how

alterations in these GPCRs contribute to tumourigenesis. GPCRs have been known for many years to regulate rapid signalling events through the generation of short-lived second messengers. However, the mechanisms by which GPCRs can sustain long-term biological responses are still being clarified. One such mechanism involves the transactivation of other signalling molecules, including the epidermal growth factor receptor (EGFR). Several distinct mechanisms for GPCR-induced EGFR transactivation have been uncovered. One pathway involves second messengerstimulated signalling, including the accumulation of intracellular Ca2+, protein kinase C activation and reactive oxygen species generation. For example, angiotensin II requires Gaq-mediated intracellular Ca²⁺ release for EGFR phosphorylation and subsequent mitogen-activated protein kinase activation [16]. GPCRs can also transactivate EGFR through the action of nonreceptor tyrosine kinases, including Src and Pyk2 [17]. This is accomplished both through direct phosphorylation of EGFR, as well as phosphorylation of adaptor proteins (including Shc) important for downstream pathway activation [18]. A third mechanism involves the GPCR-induced metalloproteinase-mediated cleavage of EGFR ligands, including proHB-EGF [19]. GPCRs can also transactivate EGFR through TACE/ADAM17-dependent transforming growth factor-α shedding, a process that requires the mitochondrial production of reactive oxygen species [20]. Direct association between GPCRs and EGFR has also been reported. In prostate cancer cells, the pro-inflammatory chemokine receptor CXCR7 associates with EGFR, and CXCR7 overexpression promotes EGFR phosphorylation [21]. Similarly, the kisspeptin receptor GPR54 interacts directly with EGFR in a ligand-dependent manner [22]. Stimulation of breast cancer cells with kisspeptin transacti-EGFR and promotes invasive behaviour. Therefore, GPCRs can transactivate growth factor signalling through a variety of mechanisms, creating opportunities for therapeutic intervention.

A recurring theme in the utilization of molecularly targeting therapeutics is the development of acquired resistance. Therefore, an intense focus is being placed on the identification of genes capable of overcoming these resistance mechanisms. To determine the contribution of GPCRs to gefitinib (EGFR inhibitor) resistance in non-small-cell lung cancer (NSCLC), a GPCR-specific microarray was used to measure GPCR expression in a resistant cell line compared to normal human lung fibroblasts [23]. Next, agonists and antagonists of differentially expressed GPCRs were tested for their ability to alter cancer cell growth. This

screening technique identified the inhibition of adenosine A2a receptors as a mechanism for therapeutic intervention of EGFR mutant NSCLC. These results suggest that various GPCRs can regulate EGFR signalling and may provide novel avenues for treatment of EGFR-driven tumours.

In addition to EGFR, cooperation between GPCRs and oncogenes or tumour suppressors has also been described in several human tumours. For example, cyclooxygenase-2-induced prostaglandin E_2 promotes colorectal tumourigenesis in the absence of functional adenomatous polyposis coli through nuclear accumulation of β -catenin [24]. Similarly, Ras upregulates prostaglandin E_2 through cyclooxygenase-2 and inhibition of the prostaglandin receptor blocks Ras-mediated transformation [25]. Therefore, a greater understanding of the interactions between GPCR and oncogene/tumour suppressor-mediated signalling may uncover novel drug targets.

GPCRs can regulate other cancer-associated signalling pathways, including signal transducer and activator of transcription-3 (STAT3) [26,27], Hippo [28,29], phosphoinositide 3-kinase (PI3K) [30] and RhoA/Rac1 [31]. The interleukin-6/Janus kinase/STAT3 signalling axis is a major mediator of cancer initiation and progression [32]. Under normal physiological conditions, this pathway is tightly regulated through negative feedback loops [33]. However, the mechanism by which STAT3 is persistently activated in cancer remained elusive. An analysis of tumour-derived myeloid cells uncovered the elevated expression of the sphingosine-1-phosphate receptor (S1PR1) (a GPCR) compared to normal splenic myeloid cells [26]. STAT3-induced S1PR1 expression promotes persistent STAT3 activation through Janus kinase 2, and S1PR1 knockdown inhibits STAT3-mediated tumourigenesis. Persistent STAT3 activation can also be accomplished through depletion of GPRC5a, a tumour suppressor in NSCLC, via the destabilization of the STAT3 inhibitor Socs3 [27]. The Hippo pathway, in which the large tumour suppressor kinases 1/2 phosphorylate and inactivate the transcriptional coactivator Yes-associated protein, controls organ size and has been implicated in human tumourigenesis [34]. Recent evidence suggests that GPCRs can both activate and inactivate large tumour suppressor kinases 1/2, depending on the coupled G-protein, in a process mediated by Rho GTPases and the actin cytoskeleton [28,29]. Activation of Rho and Rac has also been implicated in sustained proliferative signalling downstream of Gaq-linked receptors [31]. This novel pathway results in the activation of c-Jun-mediated transcription in a phospholipase C-independent manner.

GPCRs have also been implicated in the activation of PI3K, through direct binding of the catalytic p110B to the GBy subunits [30]. Disruption of this interaction can impair proliferation and invasion in phosphatase and tensin homologue-null prostate cancer cells, suggesting a possible avenue for therapeutic intervention. In uterine fibroids, GPR10 is frequently overexpressed through loss of the tumour suppressor RE1 suppressing transcription factor/neurone-restrictive silencing factor [35]. GPR10 activates PI3K/mammalian target of rapamycin signalling, and mice overexpressing GPR10 in the myometrium develop a uterine fibroid phenotype. This novel mouse model will allow the development of therapeutics for fibroids, including GPR10 antagonists. Finally, CXCR4/CXCL12 has been shown to promote growth of malignant peripheral nerve sheath tumours through activation of PI3K and β-catenin signalling [36]. Inhibition of CXCR4 activity attenuates tumourigenesis in mouse models of malignant peripheral nerve sheath tumours, providing a novel target for human disease intervention.

GPCRs regulate the Wnt and Hedgehog pathways through the receptors Frizzled and Smoothened, respectively. The role of these pathways in human tumourigenesis has been reviewed recently [37,38]. The regulation of multiple cancer-associated signalling pathways by GPCRs presents many opportunities for therapeutic intervention. In tumours with GPCR alterations, chemical inhibitors of downstream pathways may provide clinical benefit, without the need for specific GPCR antagonists. Conversely, GPCR regulation of oncogenic receptor tyrosine kinase signalling can provide a distinct mechanism for pathway inhibition. Therefore, understanding the molecular mechanisms by which GPCRs influence cancer progression is of critical importance to their utility as drug targets (Fig. 1).

GPCR regulation of hormone-sensitive tumours

A wide variety of hormones bind and activate GPCRs. Therefore, it is unsurprising that GPCRs, and their coupled G-proteins, have been shown to play critical roles in the pathogenesis of hormone responsive tumours. Among the first examples of this phenomenon was the identification of somatic mutations in the thyrotropin receptor, resulting in constitutive activation of adenylyl cyclase and hyperfunctioning thyroid adenomas [39]. Similarly, activating mutations have also been found in the luteinizing hormone receptor in Leydig-cell testicular tumours [40]. Recently, much attention has focused on the role of the novel oestrogen

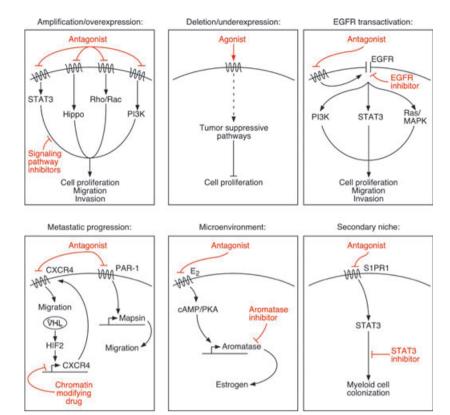


Fig. 1. GPCRs regulate multiple steps in cancer progression, including initiation, metastatic dissemination and secondary niche maintenance, through a wide array of signalling effectors. This diversity may provide novel avenues for GPCR-targeted therapeutics in the treatment of cancer. MAPK, mitogen-activated protein kinase; VHL, von Hippel–Lindau; HIF2, hypoxia-inducible factor 2; PKA, protein kinase A.

receptor G-protein coupled oestrogen receptor (GPER)-1 (formerly GPR30) in cancers of the reproductive system. It has been well established that oestrogen exerts transcriptional activation through the action of ERα and ERβ [41]. However, the molecular mechanisms by which oestrogen induced nongenomic signalling, including Ca2+ and NO generation and tyrosine kinase activation, remained largely unknown [42]. These questions were answered by the discovery that the orphan GPCR GPER-1 binds to oestrogen, resulting in intracellular Ca²⁺ accumulation and PI3K activation [43]. Recent data have begun to clarify the role of GPER-1 in oestrogen-responsive cancers. GPER-1 activation increases cancer cell proliferation from several reproductive organs, including that of the breast, endometrium, ovary and testis, and expression correlates with disease progression [44]. GPER-1 may also regulate tumour cell invasion by increasing the expression and proteolytic activity of matrix metalloproteinase-9 in ovarian cancer cells [45] and enhance fatty acid metabolism through upregulation of fatty acid synthase and EGFR transactivation [46].

Furthermore, a role for GPER-1 in regulation of the tumour microenvironment has recently been uncovered. GPER-1 is localized to the nucleus in cancer-associated fibroblasts, activates transcription, and regulates oestrogen-induced migration [47]. Therefore, GPCRs can function both at the cell surface and the nucleus to mediate cellular responses. A greater understanding of the contribution of GPER-1 to cancer pathogenesis may create opportunities for therapeutic intervention in both ER positive and negative tumours.

GPCR regulation of invasion, metastasis and the tumour microenvironment

The control of metastatic dissemination and growth at the secondary site are critical nodes in the development of novel cancer drugs. Several recent studies have highlighted the role of GPCRs in controlling cancer cell invasion and metastasis, providing targets for therapeutic intervention. In a mouse model of melanoma, the expression of endothelin receptor B enhanced spontaneous central nervous system metastases [48]. Importantly, inhibition of endothelin receptor B increased the life span in mice with visceral metastases and shrunk intracranial melanoma tumours when combined with cyclosporin A to increase drug levels in the brain parenchyma. The thrombin receptor [proteaseactivated receptor (PAR)-1] is also associated with metastatic melanoma and PAR-1 silencing can decrease tumour growth and lung metastases [49]. PAR-1 exerts these effects through transcriptional repression of the tumour suppressor Mapsin [50]. Similarly, knockdown of EGF-module containing mucinlike receptor 2 in glioblastoma cell lines reduced migration [51]. This correlates with the observation that EGF-module containing mucin-like receptor 2 is associated with poor prognosis in glioblastoma patients. In triple negative breast cancer cells, a lactoferrin/endothelin-1 pathway promotes cell invasion, and inhibition of the lactoferrin/endothelin-1 receptor blocks motility [52]. This pathway may be clinically relevant because patients with triple negative breast cancer display elevated levels of lactoferrin/endothelin-1. In kidney cancer, the von Hippel-Lindau tumour gene regulates expression suppressor chemokine receptor CXCR4, a regulator of metastatic progression, through hypoxia-inducible factor 2 [53]. Although loss of von Hippel-Lindau can initiate tumourigenesis, further epigenetic events are required for CXCR4 expression and metastatic colonization [54]. This suggests that chromatin-modifying drugs, or those targeting CXCR4, may inhibit kidney cancer progression. GPCRs can also modulate cell invasive behaviour by regulating expression of matrix metalloproteinases [55], and activation of Rho GTPases [56-581.

Preparation and maintenance of the secondary niche are critical steps during metastatic progression and both processes are regulated by GPCRs. The S1PR1-STAT3 axis supports myeloid cell colonization at the premetastatic niche, providing factors necessary for tumour cell growth [59]. Furthermore, STAT3 activity is elevated in tumour-free lymph nodes from cancer patients, suggesting that pathway inhibition may disrupt the ability of tumour cells to grow at secondary sites. In metastatic breast cancer, production of tenascin C promotes lung metastasis through the expression of the Wnt target gene LGR5, and knockdown of LGR5 decreased the ability of breast cancer cells to colonize the lung [60]. The chemokines CXCL1 and CXCL2 are amplified in invasive breast cancer and recruit myeloid cells to the primary tumour, which in turn provide factors required for cell survival and invasion [61]. This axis is stimulated by chemotherapy, providing a mechanism for chemoresistance. However, combining chemotherapy with CXCR2 inhibition in mice with lung metastases reduced metastatic burden, providing preclinical evidence for the utility of CXCR2 inhibition in the clinic. GPCRs can also regulate the metastatic switch through disruption of cell adhesion. Sphingosine 1-phosphate expression underlies the progression of T-lymphoblastic lymphoma to acute T-lymphoblastic leukaemia [62]. Increased sphingosine 1-phosphate signalling promotes cell adhesion and inhibits intravasation and dissemination in T-lymphoblastic lymphoma, whereas a selective sphingosine 1-phosphate antagonist promotes intravasation *in vivo*. GPCRs can regulate lymphatic vessel dilation and metastatic progression via cross-talk between the lymphangiogenic growth factor vascular EGF-D and prostaglandins. Finally, several GPCRs regulate tumour-associated angiogenesis, including PAR-2 [63], vGPCR [64], brain-specific angiogenesis inhibitor 1 [65] and GPR56 [66].

Because tumours are highly pro-inflammatory, regulation of the tumour microenvironment has become an important target for therapeutic intervention. GPCRs, most specifically the inflammatory chemokine receptors, control recruitment and activity of leukocytes within the tumour microenvironment and provide a means for regulating tumour-stroma interactions. Excitingly, several chemokine receptor antagonists are in clinical development for inflammatory diseases and may be co-opted for use in cancer treatment. However, because immune cells within the tumour stroma can play both positive and negative roles in tumour progression, novel inhibitors must be first tested in the appropriate cancer models. Two such studies have provided evidence that inhibition of CXCR2 may be of clinical utility in both inflammation-driven and spontaneous skin, intestinal and pancreatic tumourigenesis [67,68]. In oral squamous cell carcinoma, CXCL13/ CXCR5 signalling can activate the osteoclastogenic factor receptor activator of nuclear factor-κB ligand in stromal cells through c-Myc, resulting in bone invasion [69]. GPCRs have also been shown to regulate resistance to chemotherapy through effects on immune cell infiltration [70]. Upon doxorubicin treatment, myeloid cells infiltrate the tumour in a CCR2-dependent manner, and CCR2 null mice showed enhanced response to doxorubicin or cisplatin [70]. Therefore, these signalling axes may represent promising therapeutic targets.

GPCR signalling also provides a link between obesity and inflammation in breast cancer because the proinflammatory prostaglandin E₂ regulates aromatase expression, the enzyme necessary for oestrogen synthesis, through the cAMP/protein kinase A pathway [71]. Because elevated aromatase activity is known to increase the risk for post-menopausal hormone receptor positive breast cancer, strategies aiming to reduce inflammation may reduce the risk of breast cancer. Collectively, these studies highlight the importance of studying GPCR function in many contexts beyond the regulation of cell proliferation. Although more difficult to study in a high-throughput manner, effects on cell

migration and invasion, as well as immune function and secondary niche formation, must be taken into account when probing the cancer-associated activity of a GPCR. Furthermore, these results suggest that drugs targeting GPCRs may have utility against metastatic progression, which comprises the deadliest aspect of human cancer.

Implications and future directions

The advent of targeted therapeutics in cancer has begun to lessen our reliance on nonspecific chemotherapy. Large-scale genomic analyses of tumours have provided potential novel targets for targeted therapeutic interventions; however, separating the tumour-initiators from bystanders remains a critical challenge for both bioinformaticans and those at the bench. It is this biological validation that will truly expand the pool of targeted therapies in the clinic. It is apparent that targeted therapies will be beset by multiple mechanisms of resistance. Therefore, finding novel druggable targets is of critical importance. The GPCRs represent a class of highly druggable, yet understudied molecules in cancer. As transmembrane proteins, GPCRs can be targeted by antibodies, small molecules and peptides, neatly obviating the drug development issues associated with crossing the cell membrane. Furthermore, GPCR signalling can be modulated by agonists, antagonists and inverse agonists, allowing precise control over signalling pathways. Importantly, drug companies have extensive experience in developing GPCR-based therapeutics for a wide range of diseases, providing a knowledge base for novel target development.

An analysis of GPCR signalling in cancer reveals several emerging concepts. Advances in sequencing technologies have recently uncovered widespread GPCR alterations in human cancer [9]. However, the functional outcome of these changes remains largely unexplored. GPCRs are known to regulate multiple aspects of tumourigenesis; both tumour cell-intrinsic (proliferation, migration) and extrinsic (regulation of the tumour microenvironment, metastatic niche). Therefore, it is crucial that functional assays take into account these diverse biological processes. Furthermore, it has become evident that GPCRs can regulate cancer-associated signalling networks, resulting in pathway activation and acquired resistance. A deeper understanding of this molecular cross-talk will expand the potential for GPCR-targeted therapeutics. Advances in drug design have introduced the concept of biased ligands, which selectively regulate signalling pathways downstream of GPCRs. For these novel compounds to reach their potential, we must identify the multitude of possible signalling events triggered upon GPCR activation, as well as develop robust high-throughout assays to monitor signalling output. As the sequencing of the human genome provided the raw material for functional genomic annotation, major tumour sequencing efforts have begun to provide a comprehensive view of human cancer. We must now use this information to intelligently select and validate targets for translation of these data into real clinical advances in cancer treatment.

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References

- 1 Morris AJ & Malbon CC (1999) Physiological regulation of G protein-linked signaling. *Physiol Rev* 79, 1373–1430.
- 2 Lappano R & Maggiolini M (2011) G protein-coupled receptors: novel targets for drug discovery in cancer. *Nat Rev Drug Discov* 10, 47–60.
- 3 Young D, Waitches G, Birchmeier C, Fasano O & Wigler M (1986) Isolation and characterization of a new cellular oncogene encoding a protein with multiple potential transmembrane domains. *Cell* **45**, 711–719.
- 4 Kan Z, Jaiswal BS, Stinson J, Janakiraman V, Bhatt D, Stern HM, Yue P, Haverty PM, Bourgon R, Zheng J *et al.* (2010) Diverse somatic mutation patterns and pathway alterations in human cancers. *Nature* **466**, 869–873.
- 5 Hammerman PS, Hayes DN, Wilkerson MD, Schultz N, Bose R, Chu A, Collisson EA, Cope L, Creighton CJ, Getz G *et al.* (2012) Comprehensive genomic characterization of squamous cell lung cancers. *Nature* **489**, 519–525.
- 6 De Carvalho DD, Sharma S, You JS, Su SF, Taberlay PC, Kelly TK, Yang X, Liang G & Jones PA (2012) DNA methylation screening identifies driver epigenetic events of cancer cell survival. *Cancer Cell* 21, 655–667.
- 7 Dorsam RT & Gutkind JS (2007) G-protein-coupled receptors and cancer. *Nat Rev Cancer* 7, 79–94.
- 8 Garraway LA & Lander ES (2013) Lessons from the cancer genome. *Cell* **153**, 17–37.
- 9 O'Hayre M, Vazquez-Prado J, Kufareva I, Stawiski EW, Handel TM, Seshagiri S & Gutkind JS (2013)

- The emerging mutational landscape of G proteins and G-protein-coupled receptors in cancer. *Nat Rev Cancer* 13, 412–424.
- 10 Prickett TD, Wei X, Cardenas-Navia I, Teer JK, Lin JC, Walia V, Gartner J, Jiang J, Cherukuri PF, Molinolo A et al. (2011) Exon capture analysis of G protein-coupled receptors identifies activating mutations in GRM3 in melanoma. Nat Genet 43, 1119–1126.
- 11 Liang H, Cheung LW, Li J, Ju Z, Yu S, Stemke-Hale K, Dogruluk T, Lu Y, Liu X, Gu C *et al.* (2012) Whole-exome sequencing combined with functional genomics reveals novel candidate driver cancer genes in endometrial cancer. *Genome Res* **22**, 2120–2129.
- 12 Zhao Q, Kirkness EF, Caballero OL, Galante PA, Parmigiani RB, Edsall L, Kuan S, Ye Z, Levy S, Vasconcelos AT *et al.* (2010) Systematic detection of putative tumor suppressor genes through the combined use of exome and transcriptome sequencing. *Genome Biol* 11, R114.
- 13 Michailidou K, Hall P, Gonzalez-Neira A, Ghoussaini M, Dennis J, Milne RL, Schmidt MK, Chang-Claude J, Bojesen SE, Bolla MK et al. (2013) Large-scale genotyping identifies 41 new loci associated with breast cancer risk. Nat Genet 45, 353–361.
- 14 Garcia-Closas M, Couch FJ, Lindstrom S, Michailidou K, Schmidt MK, Brook MN, Orr N, Rhie SK, Riboli E, Feigelson HS et al. (2013) Genome-wide association studies identify four ER negative-specific breast cancer risk loci. Nat Genet 45, 392–398.
- 15 Patel MN, Halling-Brown MD, Tym JE, Workman P & Al-Lazikani B (2012) Objective assessment of cancer genes for drug discovery. *Nat Rev Drug Discov* 12, 35–50.
- 16 Eguchi S, Numaguchi K, Iwasaki H, Matsumoto T, Yamakawa T, Utsunomiya H, Motley ED, Kawakatsu H, Owada KM, Hirata Y et al. (1998) Calciumdependent epidermal growth factor receptor transactivation mediates the angiotensin II-induced mitogen-activated protein kinase activation in vascular smooth muscle cells. J Biol Chem 273, 8890–8896.
- 17 Andreev J, Galisteo ML, Kranenburg O, Logan SK, Chiu ES, Okigaki M, Cary LA, Moolenaar WH & Schlessinger J (2001) Src and Pyk2 mediate G-proteincoupled receptor activation of epidermal growth factor receptor (EGFR) but are not required for coupling to the mitogen-activated protein (MAP) kinase signaling cascade. J Biol Chem 276, 20130–20135.
- 18 Luttrell LM, Della Rocca GJ, van Biesen T, Luttrell DK & Lefkowitz RJ (1997) Gbetagamma subunits mediate Src-dependent phosphorylation of the epidermal growth factor receptor. A scaffold for G protein-coupled receptor-mediated Ras activation. *J Biol Chem* 272, 4637–4644.
- 19 Prenzel N, Zwick E, Daub H, Leserer M, Abraham R, Wallasch C & Ullrich A (1999) EGF receptor

- transactivation by G-protein-coupled receptors requires metalloproteinase cleavage of proHB-EGF. *Nature* **402**,
- 20 Myers TJ, Brennaman LH, Stevenson M, Higashiyama S, Russell WE, Lee DC & Sunnarborg SW (2009) Mitochondrial reactive oxygen species mediate GPCRinduced TACE/ADAM17-dependent transforming growth factor-alpha shedding. *Mol Biol Cell* 20, 5236–5249.
- 21 Singh RK & Lokeshwar BL (2011) The IL-8-regulated chemokine receptor CXCR7 stimulates EGFR signaling to promote prostate cancer growth. *Cancer Res* 71, 3268–3277.
- 22 Zajac M, Law J, Cvetkovic DD, Pampillo M, McColl L, Pape C, Di Guglielmo GM, Postovit LM, Babwah AV & Bhattacharya M (2011) GPR54 (KISS1R) transactivates EGFR to promote breast cancer cell invasiveness. *PLoS ONE* 6, e21599.
- 23 Kuzumaki N, Suzuki A, Narita M, Hosoya T, Nagasawa A, Imai S, Yamamizu K, Morita H, Suzuki T, Okada Y et al. (2012) Multiple analyses of Gprotein coupled receptor (GPCR) expression in the development of gefitinib-resistance in transforming nonsmall-cell lung cancer. PLoS ONE 7, e44368.
- 24 Liu X, Lazenby AJ & Siegal GP (2006) Signal transduction cross-talk during colorectal tumorigenesis. Adv Anat Pathol 13, 270–274.
- 25 Repasky GA, Zhou Y, Morita S & Der CJ (2007) Ras-mediated intestinal epithelial cell transformation requires cyclooxygenase-2-induced prostaglandin E2 signaling. *Mol Carcinog* 46, 958–970.
- 26 Lee H, Deng J, Kujawski M, Yang C, Liu Y, Herrmann A, Kortylewski M, Horne D, Somlo G, Forman S et al. (2010) STAT3-induced S1PR1 expression is crucial for persistent STAT3 activation in tumors. Nat Med 16, 1421–1428.
- 27 Chen Y, Deng J, Fujimoto J, Kadara H, Men T, Lotan D & Lotan R (2010) Gprc5a deletion enhances the transformed phenotype in normal and malignant lung epithelial cells by eliciting persistent Stat3 signaling induced by autocrine leukemia inhibitory factor. *Cancer Res* 70, 8917–8926.
- 28 Yu FX, Zhao B, Panupinthu N, Jewell JL, Lian I, Wang LH, Zhao J, Yuan H, Tumaneng K, Li H et al. (2012) Regulation of the Hippo-YAP pathway by G-protein-coupled receptor signaling. Cell 150, 780–791.
- 29 Mo JS, Yu FX, Gong R, Brown JH & Guan KL (2012) Regulation of the Hippo-YAP pathway by protease-activated receptors (PARs). *Genes Dev* 26, 2138–2143.
- 30 Dbouk HA, Vadas O, Shymanets A, Burke JE, Salamon RS, Khalil BD, Barrett MO, Waldo GL, Surve C, Hsueh C *et al.* (2012) G Protein-coupled receptor-mediated activation of p110beta by

- gbetagamma is required for cellular transformation and invasiveness. Sci Signal 5, ra89.
- 31 Vaque JP, Dorsam RT, Feng X, Iglesias-Bartolome R, Forsthoefel DJ, Chen Q, Debant A, Seeger MA, Ksander BR, Teramoto H *et al.* (2013) A genome-wide RNAi screen reveals a trio-regulated Rho GTPase circuitry transducing mitogenic signals initiated by G protein-coupled receptors. *Mol Cell* **49**, 94–108.
- 32 Yu H & Jove R (2004) The STATs of cancer new molecular targets come of age. *Nat Rev Cancer* **4**, 97–105.
- 33 Yoshimura A, Naka T & Kubo M (2007) SOCS proteins, cytokine signalling and immune regulation. *Nat Rev Immunol* **7**, 454–465.
- 34 Dong J, Feldmann G, Huang J, Wu S, Zhang N, Comerford SA, Gayyed MF, Anders RA, Maitra A & Pan D (2007) Elucidation of a universal size-control mechanism in *Drosophila* and mammals. *Cell* 130, 1120–1133.
- 35 Varghese BV, Koohestani F, McWilliams M, Colvin A, Gunewardena S, Kinsey WH, Nowak RA, Nothnick WB & Chennathukuzhi VM (2013) Loss of the repressor REST in uterine fibroids promotes aberrant G protein-coupled receptor 10 expression and activates mammalian target of rapamycin pathway. *Proc Natl Acad Sci U S A* 110, 2187–2192.
- 36 Mo W, Chen J, Patel A, Zhang L, Chau V, Li Y, Cho W, Lim K, Xu J, Lazar AJ et al. (2013) CXCR4/CXCL12 Mediate autocrine cell-cycle progression in NF1-associated malignant peripheral nerve sheath tumors. Cell 152, 1077–1090.
- 37 Anastas JN & Moon RT (2012) WNT signalling pathways as therapeutic targets in cancer. *Nat Rev Cancer* **13**, 11–26.
- 38 Kar S, Deb M, Sengupta D, Shilpi A, Bhutia SK & Patra SK (2012) Intricacies of hedgehog signaling pathways: a perspective in tumorigenesis. *Exp Cell Res* **318**, 1959–1972.
- 39 Parma J, Duprez L, Van Sande J, Cochaux P, Gervy C, Mockel J, Dumont J & Vassart G (1993) Somatic mutations in the thyrotropin receptor gene cause hyperfunctioning thyroid adenomas. *Nature* 365, 649–651.
- 40 Liu G, Duranteau L, Carel JC, Monroe J, Doyle DA & Shenker A (1999) Leydig-cell tumors caused by an activating mutation of the gene encoding the luteinizing hormone receptor. N Engl J Med 341, 1731–1736.
- 41 Hall JM, Couse JF & Korach KS (2001) The multifaceted mechanisms of estradiol and estrogen receptor signaling. *J Biol Chem* **276**, 36869–36872.
- 42 Kelly MJ & Levin ER (2001) Rapid actions of plasma membrane estrogen receptors. *Trends Endocrinol Metab* 12, 152–156.
- 43 Revankar CM, Cimino DF, Sklar LA, Arterburn JB & Prossnitz ER (2005) A transmembrane intracellular

- estrogen receptor mediates rapid cell signaling. *Science* **307**, 1625–1630.
- 44 Filardo EJ & Thomas P (2012) Minireview: G protein-coupled estrogen receptor-1, GPER-1: its mechanism of action and role in female reproductive cancer, renal and vascular physiology. *Endocrinology* **153**, 2953–2962.
- 45 Yan Y, Liu H, Wen H, Jiang X, Cao X, Zhang G & Liu G (2013) The novel estrogen receptor GPER regulates the migration and invasion of ovarian cancer cells. *Mol Cell Biochem* 378, 1–7.
- 46 Santolla MF, Lappano R, De Marco P, Pupo M, Vivacqua A, Sisci D, Abonante S, Iacopetta D, Cappello AR, Dolce V *et al.* (2012) G protein-coupled estrogen receptor mediates the up-regulation of fatty acid synthase induced by 17beta-estradiol in cancer cells and cancer-associated fibroblasts. *J Biol Chem* 287, 43234–43245.
- 47 Pupo M, Vivacqua A, Perrotta I, Pisano A, Aquila S, Abonante S, Gasperi-Campani A, Pezzi V & Maggiolini M (2013) The nuclear localization signal is required for nuclear GPER translocation and function in breast cancer-associated fibroblasts (CAFs). *Mol Cell Endocrinol* 376, 23–32.
- 48 Cruz-Munoz W, Jaramillo ML, Man S, Xu P, Banville M, Collins C, Nantel A, Francia G, Morgan SS, Cranmer LD *et al.* (2012) Roles for endothelin receptor B and BCL2A1 in spontaneous CNS metastasis of melanoma. *Cancer Res* **72**, 4909–4919.
- 49 Villares GJ, Zigler M, Wang H, Melnikova VO, Wu H, Friedman R, Leslie MC, Vivas-Mejia PE, Lopez-Berestein G, Sood AK et al. (2008) Targeting melanoma growth and metastasis with systemic delivery of liposome-incorporated protease-activated receptor-1 small interfering RNA. Cancer Res 68, 9078–9086.
- 50 Villares GJ, Zigler M, Dobroff AS, Wang H, Song R, Melnikova VO, Huang L, Braeuer RR & Bar-Eli M (2011) Protease activated receptor-1 inhibits the Maspin tumor-suppressor gene to determine the melanoma metastatic phenotype. *Proc Natl Acad Sci U S A* 108, 626–631.
- 51 Rutkowski MJ, Sughrue ME, Kane AJ, Kim JM, Bloch O & Parsa AT (2011) Epidermal growth factor module-containing mucin-like receptor 2 is a newly identified adhesion G protein-coupled receptor associated with poor overall survival and an invasive phenotype in glioblastoma. *J Neurooncol* 105, 165–171.
- 52 Ha NH, Nair VS, Reddy DN, Mudvari P, Ohshiro K, Ghanta KS, Pakala SB, Li DQ, Costa L, Lipton A *et al.* (2011) Lactoferrin-endothelin-1 axis contributes to the development and invasiveness of triple-negative breast cancer phenotypes. *Cancer Res* **71**, 7259–7269.
- 53 Staller P, Sulitkova J, Lisztwan J, Moch H, Oakeley EJ & Krek W (2003) Chemokine receptor CXCR4 downregulated by von Hippel–Lindau tumour suppressor pVHL. *Nature* 425, 307–311.

- 54 Vanharanta S, Shu W, Brenet F, Hakimi AA, Heguy A, Viale A, Reuter VE, Hsieh JJ, Scandura JM & Massague J (2013) Epigenetic expansion of VHL-HIF signal output drives multiorgan metastasis in renal cancer. *Nat Med* 19, 50–56.
- 55 Park SY, Jeong KJ, Panupinthu N, Yu S, Lee J, Han JW, Kim JM, Lee JS, Kang J, Park CG et al. (2011) Lysophosphatidic acid augments human hepatocellular carcinoma cell invasion through LPA1 receptor and MMP-9 expression. Oncogene 30, 1351–1359.
- 56 Castellone RD, Leffler NR, Dong L & Yang LV (2011) Inhibition of tumor cell migration and metastasis by the proton-sensing GPR4 receptor. *Cancer Lett* 312, 197–208.
- 57 Nithipatikom K, Gomez-Granados AD, Tang AT, Pfeiffer AW, Williams CL & Campbell WB (2012) Cannabinoid receptor type 1 (CB1) activation inhibits small GTPase RhoA activity and regulates motility of prostate carcinoma cells. *Endocrinology* 153, 29–41.
- 58 Ward Y, Lake R, Yin JJ, Heger CD, Raffeld M, Goldsmith PK, Merino M & Kelly K (2011) LPA receptor heterodimerizes with CD97 to amplify LPA-initiated RHO-dependent signaling and invasion in prostate cancer cells. *Cancer Res* **71**, 7301–7311.
- 59 Deng J, Liu Y, Lee H, Herrmann A, Zhang W, Zhang C, Shen S, Priceman SJ, Kujawski M, Pal SK et al. (2012) S1PR1-STAT3 signaling is crucial for myeloid cell colonization at future metastatic sites. *Cancer Cell* 21, 642–654.
- 60 Oskarsson T, Acharyya S, Zhang XH, Vanharanta S, Tavazoie SF, Morris PG, Downey RJ, Manova-Todorova K, Brogi E & Massague J (2011) Breast cancer cells produce tenascin C as a metastatic niche component to colonize the lungs. *Nat Med* 17, 867–874.
- 61 Acharyya S, Oskarsson T, Vanharanta S, Malladi S, Kim J, Morris PG, Manova-Todorova K, Leversha M, Hogg N, Seshan VE et al. (2012) A CXCL1 paracrine network links cancer chemoresistance and metastasis. Cell 150, 165–178.
- 62 Feng H, Stachura DL, White RM, Gutierrez A, Zhang L, Sanda T, Jette CA, Testa JR, Neuberg DS, Langenau DM *et al.* (2010) T-lymphoblastic lymphoma cells express high levels of BCL2, S1P1, and ICAM1, leading to a blockade of tumor cell intravasation. *Cancer Cell* 18, 353–366.
- 63 Yang L, Chen G, Mohanty S, Scott G, Fazal F, Rahman A, Begum S, Hynes RO & Xu L (2011) GPR56 Regulates VEGF production and angiogenesis during melanoma progression. *Cancer Res* 71, 5558–5568.

- 64 Svensson KJ, Kucharzewska P, Christianson HC, Skold S, Lofstedt T, Johansson MC, Morgelin M, Bengzon J, Ruf W & Belting M (2011) Hypoxia triggers a proangiogenic pathway involving cancer cell microvesicles and PAR-2-mediated heparin-binding EGF signaling in endothelial cells. *Proc Natl Acad Sci U S A* 108, 13147–13152.
- 65 Ma T, Jham BC, Hu J, Friedman ER, Basile JR, Molinolo A, Sodhi A & Montaner S (2010) Viral G protein-coupled receptor up-regulates angiopoietin-like 4 promoting angiogenesis and vascular permeability in Kaposi's sarcoma. *Proc Natl Acad Sci U S A* 107, 14363–14368.
- 66 Zhu D, Hunter SB, Vertino PM & Van Meir EG (2011) Overexpression of MBD2 in glioblastoma maintains epigenetic silencing and inhibits the antiangiogenic function of the tumor suppressor gene BAI1. *Cancer Res* 71, 5859–5870.
- 67 Ijichi H, Chytil A, Gorska AE, Aakre ME, Bierie B, Tada M, Mohri D, Miyabayashi K, Asaoka Y, Maeda S *et al.* (2011) Inhibiting Cxcr2 disrupts tumor-stromal interactions and improves survival in a mouse model of pancreatic ductal adenocarcinoma. *J Clin Invest* 121, 4106–4117.
- 68 Jamieson T, Clarke M, Steele CW, Samuel MS, Neumann J, Jung A, Huels D, Olson MF, Das S, Nibbs RJ *et al.* (2012) Inhibition of CXCR2 profoundly suppresses inflammation-driven and spontaneous tumorigenesis. *J Clin Invest* **122**, 3127–3144.
- 69 Sambandam Y, Sundaram K, Liu A, Kirkwood KL, Ries WL & Reddy SV (2013) CXCL13 activation of c-Myc induces RANK ligand expression in stromal/ preosteoblast cells in the oral squamous cell carcinoma tumor-bone microenvironment. *Oncogene* 32, 97–105.
- 70 Nakasone ES, Askautrud HA, Kees T, Park JH, Plaks V, Ewald AJ, Fein M, Rasch MG, Tan YX, Qiu J et al. (2012) Imaging tumor–stroma interactions during chemotherapy reveals contributions of the microenvironment to resistance. Cancer Cell 21, 488–503.
- 71 Subbaramaiah K, Morris PG, Zhou XK, Morrow M, Du B, Giri D, Kopelovich L, Hudis CA & Dannenberg AJ (2012) Increased levels of COX-2 and prostaglandin E2 contribute to elevated aromatase expression in inflamed breast tissue of obese women. *Cancer Discov* 2, 356–365.